



Report for the genetic diversity at inter- and intra-populational level

DELIVERABLE 1.6

PulpIng

Developing of Pumpkin Pulp Formulation using a Sustainable Integrated Strategy





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1. Scope of the document

The scope of this document is to capture the genetic diversity at inter- and intrapopulation level among the study pumpkin landrace. The studied material included the landraces cultivated in Greece. Following genotypic evaluation in terms of agronomic performance and determination of morpho-agronomic diversity, the most promising genotypes were molecularly characterized in order to determine the existing genetic diversity within and among genotypes under study. Molecular analysis was conducted using RAPD markers.

2. Methodology

Genetic material

The genetic material consisted of two commercial varieties and eight local landraces. Genotypes under study are listed in Table 1.

Seeds were sown in pots containing soil:perlite mixture (3:1) under controlled conditions (temperature: 25°C, photoperiod: 16h light / 8h dark). At the stage of 4 true leaves, young and healthy leaves were harvested and stored at -80 °C deep freezer until use in molecular marker analyses. For each genotype under study, four replications were used, each consisting of two true leaves.

N.	Type*	Description	Designation
1	Commercial variety 1	FYTRO FS 243	P1
2	Commercial variety 1	BIG MAX	P2
3	Local landrace 1	"Pumpkin for pie"	P3
4	Local landrace 2	"Nyxaki"	P4
5	Local landrace 3	"Melitis-BI / White"	P5
6	Local landrace 4	"Round / Deep orange"	P6
7	Local landrace 5	"Neapoli / Oval, small"	P7
8	Local landrace 6	"Lakonia / Bottle shape"	P8
9	Local landrace 7	"Makedonika / Green"	Р9
10	Local landrace 8	"K-7"	P10

Table 1. Pumpkin germplasm employed in molecular marker analysis to determine the existing genetic diversity within and among populations.

*The cultivars under study are thereof referred to as populations (P1 - P10).

DNA extraction

DNA was extracted using the CTAB method. Briefly, 200 mg of leaf tissue were placed in eppendorf tubes and grounded with 200 μ l CTAB buffer, that was previously placed in water bath (60° C) for 15 min. Subsequently, 25 mg PVP were added and samples were thoroughly mixed. Following the addition of 200 μ l CTAB, samples were thoroughly mixed and placed in a 25 °C water bath for 25 minutes. After thorough mixing (vortex), an equal volume of phenol/chloroform (200 μ l each) was added and samples were mixed to homogenization. Samples were subsequently centrifuged at 13000 rpm for 5 min, to



separate the phases, and the aqueous upper phase was transferred to a new eppendorf tube. Following the addition of 200 μ l NaCl 5M, the samples were mixed and 800 μ l of ethanol 95 % were added. Samples were mixed and placed at -20 °C overnight to precipitate the DNA. Samples were centrifuged at 13000 rpm for 10 min and the supernatant was discarded without disturbing the pellet. A second purification step was performed, using 1 ml of ice cold 70 % ethanol, and samples were centrifuged at 13000 rpm for 5 min. Ethanol was decanted and the residual ethanol was removed by drying at room temperature for 15 min. Finally, the DNA pellet was dissolved in 50 μ l mQ H₂O.

DNA concentration, in ng/ μ L, was measured using a Nanodrop spectrophotometer at 260 nm. Sample purity was estimated using the ratio 260/280, which corresponds to the absorbance at the wavelengths 260 and 280 nm (Table 2). The 260/280 ratio provides an indication of how pure the sample is from contaminating protein, with an optimal 260/280 ratio for DNA ranging to 1.80.

Sample	Genotyne	Absorbance at 260	260/280 Ratio
Sampie	Genotype	nm	200/200 Katio
1		96,7	2,16
2	Commercial variety	176,7	2,1
3	FYTRO FS 243	83,4	2,07
4		190,1	2,02
5		2.173,2	2,05
6	Commercial variety	1.717,2	2,02
7	BIG MAX	1.379,1	2,08
8		2.923,2	2,07
9		10,9	1,18
10	Local landrace 1	33,8	2,06
11	"Pumpkin for pie"	11	2,17
12		10,6	2,14
13		2.943,2	2,09
14	Local landrace 2	2.005	2,07
15	"Nyxaki"	2.171,2	2,09
16		1.266,4	2,12
17		409	2,08
18	Local landrace 3	217,4	2,14
19	"Melitis-BI / White"	331,2	2,03
20		184,2	1,84
21	Legal landrage 4	317,4	2,04
22	"Dourd / Door	98,4	1,99
23	Kound / Deep	773,7	1,88
24	orange	243,8	2,14
25	Local landroop 5	66	2,16
26	Local landrace 3	231,4	2,11
27	ineapoii / Oval,	290,3	2,11
28	sman	241,7	2,01
29	Local los duces (99,5	2,07
30	Local landrace o	87,6	2,13

Table 2. Nanodrop readings for genomic DNA of pumpkin samples under study.



31	"Lakonia / Bottle	86,5	2,02
32	shape"	96,4	2,17
33		92,4	2,16
34	Local landrace 7	112,8	1,83
35	"Makedonika / Green"	105,6	2,07
36		142,1	2,01
37		3.972,2	2,03
38	Local landrace 8	3.553,9	2,05
39	"K-7"	3.016,9	2,08
40		3.642,2	2,06

RAPD analysis

For the RAPD analysis, random 10-mer primers were used for amplification, using the DNA from four individual plants of each genotype. PCR reaction and amplification conditions were accordingly standardized. Primers used for RAPD analyses are listed in Table 3, while the PCR reaction conditions are described in Table 4.

Following RAPD-PCR reactions, the amplification products were separated in 1.5-2 % agarose gels and detected by staining with MIDORI Green Advance, which is a non-carcinogenic and less mutagenic dye routinely employed as an alternative to the traditional ethidium bromide-mediated staining of nucleic acids. PCR products were visualized under UV light.

N.	Туре	Primer Name	Sequence 5' \rightarrow 3'	Citation
1	RAPD	OPA-04	AATCGGGCTG	Radwan, 2014. Molecular discrimination
2	RAPD	OPB-01	GTTTCGCTCC	and genetic relationships between some
3	RAPD	OPB-02	TGATCCCTGG	cultivars of C. pepo using RAPD analysis.
4	RAPD	OPB-04	GGACTGGAGT	African Journal of Biotechnology, 11(3):
5	RAPD	OPG-02	GGCACTGAGG	1202-1209.
6	RAPD	OPZ-03	CAGCACCGCA	
7	RAPD	CB9	GGTGACGCAG	Ntuli et al., 2015. Genetic diversity in
8	RAPD	CB12	AGTCGACGCC	Cucurbita pepo landraces revealed by
9	RAPD	CB13	ACGCATCGGA	RAPD and SSR markers. Scientia
10	RAPD	CB15	GGCTGGTTCC	Horticulturae, 189: 192-200.
11	RAPD	CB17	GTAACCAGCC	

Table 3. Nucleotide sequences of primers used for molecular marker analysis.

Table 4. PCR reaction conditions used in RAPD marker analyses.

PCR Reaction Mix									
RAPD Primers		Reagent	Final	Volume					
			concentrat	ion					
OPA-04	Buffer		1x	5 µL					



OPB-01	MgCl ₂	1.5 mM	1.5 μL
OPB-02	RAPD primer	10 pmole	1 μL
OPB-04	dNTPs	200 µM each	0.5 μL
OPG-02	Taq polymerase (KAPA Taq DNA	0.5 u	0.2 µL
	Polymerase)		·
OPZ-03	DNA	50 ng	2.5 μL
	ddH ₂ O	-	14.3 μL
	Total volume		25 μĹ
	PCR Program		· · · · · · · · · · · · · · · · · · ·
95 °C	3 min		
95 °C	2 min		
37 °C	1 min	45 cyc	eles
72 °C	2 min		
72 °C	10 min		
4 °C	overnight		
	Gel Electrophoresis		
Agarose gel: 1.5	-2 % TBE	Ladder: 100 bp DN	A Ladder (NEB)

PCR Reaction Mix								
RAPD Primers	Reagent	Final	Volume					
		concentration						
CB9	Buffer	1x	5µl					
CB12	MgCl ₂	1.5 mM	1.5 μL					
CB13	RAPD primer	0.4 µM	1 µĹ					
CB15	dNTPs	200 µM each	0.5 μL					
CB17	Taq polymerase (GoTaq Master Mix)	0.5 u	0.2 µL					
	DNA	50 ng	2.5 µL					
	H ₂ O	-	14.3 μL					
	Total volume		25 µL					
	PCR Program							
95 °C	5 min							
95 °C	30 sec							
36 °C	30 sec	40 c	cycles					
72 °C	1 min		-					
72 °C	4 min							
4 °C	overnight							
	Gel Electrophoresis	5						
Agarose gel: 1.5 -	- 2 % TBA	Ladder: 100 bp D	NA Ladder (NEB)					

<u>Data analysis</u>

Each PCR reaction was run in duplicate and only discrete and reproducible bands were analyzed. RAPD gel images were analyzed using GelAnalyzer 23.1 software (<u>http://www.gelanalyzer.com/</u>). The band size was determined using a 100 bp DNA Ladder (NEB).

The RAPD profiles for each primer were scored for presence or absence, as a binary character, and transferred to a binary data matrix using MS Excel. The presence coded



"1" and absence coded "0" of each band in a data matrix. Monomorphic bands were excluded from the analysis. An example data matrix using RAPD marker A04 is presented in the Appendix (Table 1). The total number of bands, number of polymorphic bands, and percentage of polymorphic bands were counted.

The binary matrix that included 40 individuals and 215 generated RAPD loci was used for further data analysis. The genetic diversity indices were calculated using the GenAlEx 6.5 package. The analysis involved determination of the observed allele number (N), the number of different alleles (Na), the effective allele number (Ne), Shannon's information index (I), Nei's genetic distance (D) and genetic identity (IN), gene diversity (h), and unbiased gene diversity (uh). The formulas employed for the determination of genetic parameters under study are presented in the Appendix (Table2).

Distribution of the genetic variation within populations and among populations based on the RAPD marker profiles was determined by analysis of molecular variance (AMOVA). Further, the pairwise population differentiation (PhiPT) was calculated with random permutation number N = 999.

Results

RAPD marker polymorphism

In the framework of characterizing the set of local landraces collected from various cultivation areas in Greece, RAPD marker analysis was employed. Eleven (11) primers were used to estimate the genetic diversity between two commercial varieties and eight local landraces by amplifying the extracted DNA using RAPD-PCR analysis.

The RAPD profiles generated by the eleven RAPD primers A-04, B-01, B-02, B-04, G-02, Z-03, CB-09, CB-12, CB-13, CB-15 and CB-19 are illustrated in Figures 1-11, respectively. The RAPD primer B-02 did not yield discrete and reproducible bands and, as such, was excluded from the analysis.





Figure 1. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker OPA-04. Ladder: 100 bp DNA Ladder (NEB).



Figure 2. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker OPB-01. Ladder: 100 bp DNA Ladder (NEB).





Figure 3. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker OPB-02. Ladder: 100 bp DNA Ladder (NEB).



Figure 4. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker OPB-04. Ladder: 100 bp DNA Ladder (NEB).







Figure 5. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker OPG-02. Ladder: 100 bp DNA Ladder (NEB).



Figure 6. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker OPZ-03. Ladder: 100 bp DNA Ladder (NEB).







Figure 7. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker CB-9. Ladder: 100 bp DNA Ladder (NEB).



Figure 8. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker CB-12. Ladder: 100 bp DNA Ladder (NEB).





Figure 9. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker CB-13. Ladder: 100 bp DNA Ladder (NEB).



Figure 10. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker CB-15. Ladder: 100 bp DNA Ladder (NEB).



	FYTRO-FS243				_	BIG M	MAX	_	-	Land	race	1		andr	ace 2	_		Land	race 3	3
L	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20
-																				
	-			_																
-																				
						=	=	=				=	=	-	=	=		-	=	=
=																				
=																				
																				_
	_	Land	race	4		Landr	ace 5	<u>;</u>	1	andr	ace (5	_	andr	ace 7		_	andı	race 8	3
L	21	Land 22	race 23	4 24	25	Landr 26	ace 5	; 28	 29	andr 30	ace 6 31	5 32	33	andr 34	ace 7 35	36	<u> </u> 37	andı 38	race 8 39	<u>3</u> 40
L	21	Land 22	race 23	4 24	25	Landr 26	ace 5 27	28	29	andr 30	ace (31	5 32	33	andr 34	ace 7 35	36	37	<u>andı</u> 38	ace 8 39	<u>3</u> 40
L	21	Land 22	race 23	<u>4</u> 24	25	Landr 26	ace 5 27	; 28	 29	<u>andr</u> 30	ace (31	32	33	<u>andr</u> 34	ace 7 35	36	<u> </u> 37	<u>andı</u> 38	race 8 39	<u>3</u> 40
L	21	Land 22	race 23	4 24	25	Landr 26	ace 5 27	28	29	<u>andr</u> 30	ace (31	32	33	<u>andr</u> 34	ace 7 35	36	37	<u>andı</u> 38	race 8 39	<u>3</u> 40
L	21	Land 22	race 23	24	25	Landr 26	race 5 27	28	29	<u>andr</u> 30	ace (31	32	33	andr 34	ace 7 35	36	37	<u>andı</u> 38	race 8 39	3 40
	21	Land 22	race 23	<u>4</u> 24	25	Landr 26	27	28	29	andr 30	ace (31	32	33	andr 34	ace 7 35	36	37	<u>Landı</u> 38	ace (39	3 40
	21	Land 22	race 23	4 24	25	Landr 26	ace 5 27	28	29	andr 30	ace (31	32	33	andr 34	ace 7 35	36	37	<u>andı</u> 38	ace (39	3 40
	21	Land 22	race 23	4 24	25	Landr 26	27	28	29	andr 30	ace (32	33	andr 34	ace 7	36	37	Landı 38	ace 8 39	3 40

Figure 11. RAPD profiles illustrating bands amplification in the set of populations of *C. pepo* using the RAPD marker CB-17. Ladder: 100 bp DNA Ladder (NEB).

The set of 10-decamer arbitrary RAPD primers employed yielded highly polymorphic patterns and generated a total number of 215 amplification products. The degree of polymorphism and the size of amplification products are presented in Table 5.

RAPD	Sequence (5' –	NTD	NDD	Dand size range (hn)	DDD (0/)
primer	3')	NID	INF D	Danu size range (up)	FFD (70)
OPA-04	AATCGGGGCTG	11	11	240 - 1400	100
OPB-01	GTTTCGCTCC	10	10	166 - 1190	100
OPB-04	GGACTGGAGT	21	21	209 -1946	100
OPG-02	GGCACTGAGG	27	27	227 - 2618	100
OPZ-03	CAGCACCGCA	21	21	256 - 2975	100
CB-09	GGTGACGCAG	23	23	211 -2399	100
CB-12	AGTCGACGCC	28	28	318 - 2406	100
CB-13	ACGCATCGGA	48	47	176 - 2879	97,91
CB-15	GGCTGGTTCC	26	26	351 - 2925	100

Table 5. Analysis of the RAPD profiles of *C. pepo* populations under study. The genetic material consisted of two commercial varieties and eight local landraces.

NTB: number of total bands, NPB: number of polymorphic bands, bp: base pair, PPB: percentage of polymorphic bands.

Amplification products ranged in size from 166 - 2975 bp. The highest number of bands was obtained using the RAPD marker CB-13 (48 bands), followed by markers CB-12,



OPG-02 and CB-15 (28, 27 and 26 bands, respectively). In contrast, the lowest number of bands was recorded in marker OPB-01 and OPA-04 (10 and 11 bands, respectively). The number of polymorphic bands ranged from 10 to 47 (OPB-01 and CB-13, respectively), with a mean value of 23,78 polymorphic bands. Overall findings are indicative of the fact that the RAPD markers employed in this study are suitable for assessing the genetic diversity existing within and among populations of *C. pepo*.

RAPD	Saguanaa (5? - 2?)	Ν	Na	Ne	Ι	h	uh
primer	Sequence $(5^{\circ} - 5^{\circ})$						
OPA-04	AATCGGGGCTG	4	0.491	1.089	0.072	0.050	0.067
OPB-01	GTTTCGCTCC	4	0.660	1.186	0.160	0.109	0.148
OPB-04	GGACTGGAGT	4	0.590	1.160	0.136	0.093	0.125
OPG-02	GGCACTGAGG	4	0.541	1.120	0.102	0.070	0.094
OPZ-03	CAGCACCGCA	4	0.833	1.234	0.199	0.136	0.185
CB-09	GGTGACGCAG	4	0.774	1.195	0.160	0.110	0.150
CB-12	AGTCGACGCC	4	0.864	1.250	0.203	0.140	0.192
CB-13	ACGCATCGGA	4	0.604	1.203	0.167	0.115	0.157
CB-15	GGCTGGTTCC	4	0.704	1.210	0.177	0.121	0.163
Total*		40	0.678	1.191	0.159	0.109	0.148

Table 6. Polymorphism of different RAPD primers and genetic diversity indices within ten *C. pepo* populations under study.

*Grand mean over genetic loci and populations

N: number of samples per population, Na: number of different alleles, Ne: number of effective alleles, I: Shannon's Information Index, h: gene diversity, uh: unbiased gene diversity.

Genetic diversity analysis

The genetic diversity existing within and among populations of *C. pepo* was determined based on the polymorphic band percentage (PBP %), the number of different alleles (Na), the number of effective alleles (Ne), Shannon's Information Index (I), gene diversity, Nei's genetic distance (D) and Nei's genetic identity (IN). The results for all parameters of genetic diversity are presented in Table 7. The percentage of polymorphic loci for pumpkin populations based on data from individual marker analysis is provided in the Appendix (Table 3).

The RAPD marker analysis revealed an average polymorphic band percentage (PBP) that ranged from 18.14 to 32.09 among *C. pepo* populations under study. The highest PBP was recorded in P6 (32.09 %), followed by P7 (30.23 %), whereas the lowest PBR was found in P5 and P10 (18.14 %). The average number of different alleles (Na) ranged from 0.530 to 0.781 for P10 and P3, respectively (Table 7).

In relation to the number of effective alleles (Ne), the values ranged from 1.145 to 1.228. The highest Ne value was recorded in P6 (1.228), while P5 and P10 showed the lowest Ne (1.145). In accordance with findings related to PBP and Ne, P6 showed also the highest Shannon index (I = 0.192) and the highest gene diversity (h = 0.131).



Accordingly, the lowest respective values were recorded in P5 and P10 (I = 0.115, h = 0.081) (Table 7).

Population	Cultiver News	PBP	Ν	Na	Ne	Ι	h	uh
	Cultivar Name	(%)						
P1	FYTRO FS 243	22.79	4	0.600	1.167	0.138	0.095	0.126
P2	BIG MAX	29.77	4	0.726	1.208	0.177	0.121	0.161
P3	Pumpkin for pie	29.77	4	0.781	1.210	0.178	0.122	0.162
P4	Nyxaki	28.37	4	0.735	1.206	0.171	0.117	0.157
P5	Melitis-BI / White	18.14	4	0.535	1.145	0.115	0.081	0.121
P6	Round / Deep	32.09	4	0.753	1.228	0.192	0.131	0.175
	orange							
P7	Neapoli / Oval,	30.23	4	0.735	1.206	0.178	0.121	0.161
	small							
P8	Lakonia / Bottle	27.44	4	0.707	1.196	0.165	0.113	0.150
	shape							
P9	Makedonika / Green	25.58	4	0.679	1.198	0.158	0.110	0.147
P10	K-7	18.14	4	0.530	1.145	0.115	0.081	0.121
Total*			40	0.678	1.191	0.159	0.109	0.148

Table 7. Genetic diversity indices of *C. pepo* populations based on RAPD marker analysis.

*Grand mean over genetic loci and populations

PBP: percentage of polymorphic bands, N: number of samples, Na: number of different alleles, Ne: number of effective alleles, I: Shannon's Information Index, h: gene diversity (heterozygosity value), uh: unbiased gene diversity.

The band patterns for binary (haploid) data by populations are illustrated in Graph 1. The total band patterns for binary (haploid) data by populations for individual markers as well as for the entire set of markers are provided in the Appendix.

As shown in Graph 1, the highest number of locally common bands, present in 25 % or fewer populations, was found in P3 (10 bands) and P6 (8 bands). To the contrary, the lowest number was found in P5 (1 band). Furthermore, the analysis revealed that populations 1, 4 and 7 present the highest number of unique bands to a single populations (4 unique bands). In contrast, the lowest number of unique bands was found in populations 5, 8 and 9 (1 unique band).





Graph 1. Band patterns of RAPD markers by populations.

[No. Bands = No. of different bands, No. Bands Freq. $\geq 5\%$ = No. of Different Bands with a Frequency $\geq 5\%$,

No. Private Bands = No. of Bands Unique to a Single Population, No. LComm Bands (<=25%) = No. of Locally Common Bands (Freq. >= 5%) Found in 25% or Fewer Populations, No. LComm Bands (<=50%) = No. of Locally Common Bands (Freq. >= 5%) Found in 50% or Fewer Populations]

Further, analysis of molecular variance (AMOVA) was conducted as a means to determine the distribution of genetic diversity both among and within populations under study. AMOVA revealed that 59 % of total genetic variation was attributed to differences among populations and 41 % of genetic variance was accordingly attributed to differences within populations (Table 8). The PhiPT value was 0.587.

Table 8. Analysis of molecular variance (AMOVA) of ten *C. pepo* populations using RAPD markers. The genetic material consisted of two commercial varieties and eight local landraces.

Source of variation	DF	SS	MS	Est. Var.	%	PhiPT
Among populations	9	928,171	103,130	22,922	59%	0.587**
Within populations	30	451,750	16,134	16,134	41%	
Total	39	1379,921		39,056	100%	

DF: degrees of freedom, SS: sum of squares, MS: mean square deviations, Est.Var.: estimated variance, percentage of total variance (%), PhiPT: significance of variance (p-value) [PhiPT = AP / (WP + AP) = AP / TOT], ** p < 0.001.

The pairwise population PhiPT values were estimated in order to estimate the genetic differentiation among populations under study. The PhiPT values ranged from 0.345 to 0.671. The analysis revealed a high differentiation (PhiPT = 0.671) of P3 and P9 populations, whereas the lowest differentiation was recorded between P8 and P9 populations (PhiPT = 0.345) (Table 9).

Table 9. Pairwise population PhiPT values among ten *C. pepo* populations using RAPD markers. The values are calculated with permutation number N = 999.

Population 1	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10



P1	-									
P2	0.547	-								
P3	0.624	0.555	-							
P4	0.616	0.494	0.587	-						
P5	0.665	0.631	0.454	0.610	-					
P6	0.584	0.541	0.457	0.578	0.464	-				
P7	0.505	0.585	0.576	0.593	0.588	0.524	-			
P8	0.642	0.620	0.631	0.529	0.655	0.570	0.543	-		
P9	0.651	0.642	0.671	0.590	0.659	0.609	0.581	0.345		
P10	0.652	0.636	0.666	0.591	0.670	0.606	0.593	0.541	0.424	-

Further, Nei's genetic distance and identity were determined in order to provide estimates for the genetic relationship among populations. Nei's genetic distance ranged from 0.139 to 0.510, while the genetic identity ranged accordingly from 0.600 to 0.870 (Table 10). In relation to genetic distance among populations, the highest values were recorded for P3 and P9 (D = 0.510), whereas the corresponding lowest values were found in P8 and P9 populations (D = 0.139). Accordingly, P8 and P9 showed the highest genetic identity (IN = 0.870), followed by P9 and P10 (IN = 0.845). In contrast, P3 and P9 were the least identical populations (IN = 0.600). The Nei's genetic distance and genetic identity based on the analysis of individual markers are provided in the Appendix.

Table 10. Nei's genetic distance and genetic identity among ten *C. pepo* populations using RAPD markers. The genetic material consisted of two commercial varieties and eight local landraces.

		Pairw	vise Popu	lation M	atrix of Ne	i Genetic	Distanc	e		
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
P1	0.000									
P2	0.268	0.000								
P3	0.369	0.322	0.000							
P4	0.348	0.250	0.360	0.000						
P5	0.370	0.390	0.201	0.347	0.000					
P6	0.330	0.321	0.237	0.366	0.221	0.000				
P7	0.229	0.364	0.351	0.369	0.325	0.301	0.000			
P8	0.379	0.404	0.426	0.272	0.409	0.345	0.293	0.000		
P9	0.387	0.440	0.510	0.340	0.407	0.402	0.335	0.139	0.000	
P10	0.350	0.399	0.462	0.321	0.373	0.377	0.331	0.256	0.168	0.000



		Pairw	vise Popu	ılation M	atrix of No	ei Genetic	ldentity	ý		
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
P1	1.000									
P2	0.765	1.000								
P3	0.691	0.724	1.000							
P4	0.706	0.779	0.698	1.000						
P5	0.690	0.677	0.818	0.707	1.000					
P6	0.719	0.725	0.789	0.693	0.802	1.000				
P7	0.795	0.695	0.704	0.692	0.723	0.740	1.000			
P8	0.684	0.668	0.653	0.762	0.664	0.708	0.746	1.000		
P9	0.679	0.644	0.600	0.712	0.665	0.669	0.715	0.870	1.000	
P10	0.705	0.671	0.630	0.725	0.689	0.686	0.718	0.774	0.845	1.000

Principal coordinate analysis (PCoA) was used to generate a coordinate graph which provides visualization of the genetic relationships among populations of *C. pepo*. In the coordinate graph, the points represent the individuals from the ten populations under study (Figure 12). The results of PCoA are in good agreement with those obtained by the genetic distance and identity indices, supporting the separation of individuals into three main groups. As shown in Figure 12, populations P8, P9 and P10 are clustered together, while the analysis further revealed close genetic relationships among P3, P5 and P6. The third group, involving populations P1, P2, P4 and P7, shows relatively greater genetic divergence as compared to the others.



Figure 12. Principal coordinate analysis of ten populations of C. pepo based on RAPD molecular marker analysis.

Conclusively, the findings underline the suitability of RAPD markers for determining the genetic diversity both at the intra- and inter-populational level. RAPD analysis of the germplasm under study revealed a significant level of genetic diversity both within and among populations, while it is interesting both from an agronomic and breeding



perspective that the Greek local landraces derived from different geographic regions under study exhibit a considerable genetic variation.



APPENDIX

Table 1. A data matrix for RAPD marker A04. Polymorphic bands are scored as a binary character, "1" stands for a presence of a band and "0" stands for its absence.

Lane 7	# 1400	1300	1200	1100	970	690	630	460	380	300	240
2	1	0	0	0	0	0	0	0	0	0	0
2	0	0	0	0	0	0	l	0	0	0	0
$\frac{2}{3}$	0	0	0	0	0	0	0	0		0	0
3	$\stackrel{1}{0}$	0	0	0	0	0	1	0	0	0	ŏ
4	ĭ	ŏ	ŏ	ŏ	ŏ	ŏ	Ō	ŏ	ŏ	ŏ	ŏ
4	0	0	0	0	0	0	1	0	0	0	0
4	0	0	0	0	0	0	0	0	1	0	0
4	0	0	0	0	0	0	0	0	0	0	1
5		0	0	0	0	0	0	0	0	0	0
5	0	0	0	0	0	0	$\stackrel{1}{0}$	0	1	0	0
5	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ	$\dot{0}$	ŏ	1
6	Õ	Ō	1	Ō	Ŏ	Ō	Ŏ	Ō	Ō	Ō	Ō
6	0	0	0	1	0	0	0	0	0	0	0
6	0	0	0	0	0	0	1	0	0	0	0
6	0	0	0	0	0	0	0		0	0	0
0	0	0	0	0	0	0	0	0		0	0
7	0	0	1	0	0	0	0	0	Ő	0	$\dot{0}$
, 7	ŏ	ŏ	Ô	ĭ	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ
7	0	0	0	0	0	0	1	0	0	0	0
7	0	0	0	0	0	0	0	1	0	0	0
7	0	0	0	0	0	0	0	0	1	0	0
8	0	0		0	0	0	0	0	0	0	0
0 8	0	0	0	$ \begin{bmatrix} 1 \\ 0 \end{bmatrix} $	1	0	0	0	0	0	0
8	ŏ	ŏ	ŏ	ŏ	$\dot{0}$	1	ŏ	ŏ	ŏ	ŏ	ŏ
8	Ō	Ō	Ō	Ō	Ŏ	Ō	ĺ	Õ	Ō	Ō	Õ
8	0	0	0	0	0	0	0	0	1	0	0
8	0	0	0	0	0	0	0	0	0	0	1
9	0	0		0	0	0	0	0	0	0	0
9	0	0	0		0	0	1	0	0	0	0
9	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ	$\dot{0}$	1	ŏ	ŏ	ŏ
9	Ō	Ō	Ō	Ō	Ŏ	Ō	Ŏ	Ō	1	Ō	Õ
10	0	0	0	0	1	0	0	0	0	0	0
10	0	0	0	0	0	0	1	0	0	0	0
11 11	0	0	0	0		0	0	0	0	0	0
11	0	0	0	0	0	0	$\stackrel{1}{0}$	1	0	0	0
12	ŏ	ŏ	ŏ	ŏ	ĩ	ŏ	ŏ	$\hat{0}$	ŏ	ŏ	ŏ
12	0	0	0	0	0	0	1	0	0	0	0
13	0	0	0	0	1	0	0	0	0	0	0
13	0	0	0	0	0	0	1	0	0	0	0
14	0	0	0		0	0	0	0	0	0	0
14	0	0	0	0	$\stackrel{1}{0}$	0	1	0	0	0	0
14	ŏ	Ő	ŏ	ŏ	ŏ	ŏ	$\dot{0}$	ŏ	ŏ	1	ŏ
14	Ŏ	ŏ	Ŏ	Ŏ	Ŏ	Ŏ	ŏ	Ŏ	ŏ	Ō	ľ
15	0	0	0	1	0	0	0	0	0	0	0
15	0	0	0	0	1	0	0	0	0	0	0
15	0	0	0	0	0	0		0	0	0	0
15	0	0	0	0	0	0	0	1	0	1	0
16	Ŏ	ŏ	ŏ	1	ŏ	ŏ	ŏ	ŏ	ŏ	0	Ő
16	Ŏ	Ŏ	Ŏ	Ô	ĩ	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ





16	0	0	0	0	0	0	1	0	0	0	0
16 16	0	0	0	0	0	0	0	0	0	1	0
17	0 0	0	0	1	0	0	0	0	0	0	0
17	0	0	0	0	1	0	0	0	0	0	0
17	0	0	0	0	0	0	1	0	0	0	0
$17 \\ 17$	0	0	0	0	0	0	0		0	1	0
18	ŏ	ŏ	ŏ	ŏ	1 1	ŏ	ŏ	ŏ	ŏ	Ô	ŏ
18	0	0	0	0	0	0	1	0	0	0	0
19 10	0	0	0	0		0	0	0	0	0	0
20	ŏ	0	0	0 0	1	0	$\overset{1}{0}$	0	0 0	0 0	0
20	0	0	0	0	0	0	1	0	0	0	0
21	0	0	0	1	0	0	0	0	0	0	0
$\frac{21}{22}$	0	0	0	0	0	1	0	0	0	0	0
$\frac{22}{22}$	ŏ	0 0	0 0	0	ŏ	1	ŏ	ŏ	ŏ	ŏ	Ŏ
23	0	0	0	1	0	0	0	0	0	0	0
23	0	0	0	0	0		0	0	0	0	0
24	0	0	0		0	1	0	0	0	0	0
25	ľ	Ŏ	Ŏ	Ŏ	Ŏ	Ō	Ŏ	Ŏ	Ŏ	Ŏ	Ŏ
25	0	1	0	0	0	0	0	0	0	0	0
25	0	0	0	0	0	0		0	0	0	0
$\frac{23}{26}$	1	0	0	0	0	0	0	0	$\overset{1}{0}$	0	0
26	Ō	1	Õ	Õ	Ŏ	Õ	Ō	Ŏ	Ŏ	Õ	Õ
26	0	0	0	0	0	0	1	0	0	0	0
26 27	0	0	0	0	0	0	0	0	1	0	0
$\frac{2}{27}$	$\overset{1}{0}$	1	0	0 0	0	0	0 0	0	0	0	0
27	0	0	0	0	0	0	1	0	0	0	0
27	0	0	0	0	0	0	0	0	1	0	0
28 28		0	0	0	0	0	0	0	0	0	0
$\frac{20}{28}$	ŏ	0	ŏ	ŏ	ŏ	ŏ	ĩ	ŏ	ŏ	ŏ	Ŏ
28	0	0	0	0	0	0	0	0	1	0	0
29	0		0	0	0	0	0	0	0	0	0
29	Ő	0	$\overset{1}{0}$	1	0	0	0	0	0	0	0
2 9	Ŏ	Ŏ	Ŏ	Ō	Ŏ	ľ	Ŏ	Ŏ	Ŏ	Ŏ	Ŏ
29	0	0	0	0	0	0	0	0	1	0	0
30	0		0	0	0	0	0	0	0	0	0
30	ŏ	Ŏ	0	1	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ	0
30	0	0	0	0	0	1	0	0	0	0	0
30	0	0	0	0	0	0	0	0		0	0
31	0		1	0	0	0	0	0	0	0	0
31	Ŏ	Ŏ	Ō	Ŏ	Ŏ	ľ	Ŏ	Ŏ	Ŏ	Ŏ	Ŏ
31	0	0	0	0	0	0	0	0	1	0	0
32	0		0	0	0	0	0	0	0	0	0
32	Ő	0	$\overset{1}{0}$	1	0	0	0 0	0	0	0	0
32	0	0	0	0	0	0	0	0	1	0	0
33	0	1	0	0	0	0	0	0	0	0	0
<i>33</i>	0	0	$1 \\ 0$	0	0	1	0	0	0	0	0
33	ŏ	ŏ	ŏ	ŏ	ŏ	Ô	ŏ	ŏ	1	ŏ	ŏ
34	0	1	0	0	0	0	0	0	0	0	0
34 31	0	0		0	0	0	0	0	0	0	0
54	U	U	U	U	U	1	U	U	U	U	U



34	0	0	0	0	0	0	0	0	1	0	0
35	ŏ	ĭ	ŏ	ŏ	ŏ	ŏ	ŏ	ŏ	Ō	ŏ	ŏ
35	ŏ	Ō	ľ	Ŏ	Ŏ	ŏ	Ŏ	ŏ	ŏ	Ŏ	Ŏ
35	Ŏ	Ŏ	Ō	Ŏ	Ŏ	Ŏ	Ŏ	Ŏ	Ĩ	Ŏ	Ŏ
36	Ŏ	ľ	Ŏ	Ŏ	Ŏ	Ŏ	Ŏ	Ŏ	Ō	Ŏ	Ŏ
36	Õ	Ō	Ĩ	Õ	Õ	Õ	Õ	Õ	Õ	Õ	Õ
36	Ŏ	Ŏ	Ō	Ŏ	Õ	Ŏ	Õ	Ŏ	Ĩ	Õ	Ŏ
37	0	1	0	0	0	0	0	0	0	0	0
37	0	0	1	0	0	0	0	0	0	0	0
37	0	0	0	0	0	1	0	0	0	0	0
37	0	0	0	0	0	0	0	0	1	0	0
38	0	1	0	0	0	0	0	0	0	0	0
38	0	0	1	0	0	0	0	0	0	0	0
38	0	0	0	0	0	1	0	0	0	0	0
38	0	0	0	0	0	0	0	0	1	0	0
39	0	1	0	0	0	0	0	0	0	0	0
39	0	0	1	0	0	0	0	0	0	0	0
39	0	0	0	0	0	1	0	0	0	0	0
39	0	0	0	0	0	0	0	0	1	0	0
						4.					



Genetic parameter	Formula	Designations
Ne	Ne = No. of Effective Alleles = $1 / (p^2 + q^2)$	
I	I = Shannon's Information Index = - 1* (p * Ln (p) + q * Ln(q))	
h	$h = Diversity = 1 - (p^2 + q^2)$	Where for Haploid Binary data, $p =$ Band Freq. and $q = 1 - p$
uh	uh = Unbiased Diversity = (N / (N-1)) * h	Where for Haploid Binary data, $p =$ Band Freq. and $q = 1 - p$
PhiPT	PhiPT = AP / (WP + AP) = AP / TOT	AP = Est. Var. Among Pops WP = Est. Var. Within Pops

 Table 2. Formulas used for the estimation of genetic parameters.

Table 3. Percentage of polymorphic loci for pumpkin populations under study.

			Percer	itage of I	Polymor	рис гос	21			
Population				% Po	lymorph	ism				
	OPA-	OPB-	OPB-	OPG-	OPZ-	CB-	CB-	CB-	CB-	Total
	04	01	04	02	03	09	12	13	15	TUTAL
P1	18,18	20,00	28,57	11,11	42,86	8,70	25,00	16,67	38,46	22,79
P2	36,36	50,00	28,57	3,70	28,57	26,09	32,14	43,75	23,08	29,77
P3	18,18	40,00	47,62	44,44	42,86	13,04	7,14	25,00	38,46	29,77
P4	18,18	30,00	19,05	3,70	52,38	34,78	39,29	25,00	34,62	28,37
P5	0,00	20,00	0,00	3,70	33,33	4,35	50,00	18,75	19,23	18,14
P6	0,00	50,00	14,29	44,44	19,05	30,43	35,71	37,50	38,46	32,09
P7	0,00	10,00	28,57	22,22	42,86	34,78	39,29	29,17	38,46	30,23
P8	18,18	20,00	28,57	11,11	33,33	26,09	32,14	35,42	26,92	27,44
P9	9,09	10,00	14,29	14,81	23,81	43,48	53,57	14,58	34,62	25,58
P10	0,00	20,00	19,05	11,11	14,29	39,13	14,29	27,08	3,85	18,14
Mean	11,82	27,00	22,86	17,04	33,33	26,09	32,86	27,29	29,62	26,23
SE	3,85	4,73	4,00	4,91	3,82	4,20	4,57	2,99	3,63	1,58

Percentage of Polymorphic Loci



Total Band Patterns for Binary (Haploid) Data by Populations

OPA-04											
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
No. Bands	4	8	3	6	2	2	4	5	4	3	
No. Bands Freq. >= 5%	4	8	3	6	2	2	4	5	4	3	
No. Private Bands	0	0	0	1	0	0	0	0	0	0	
No. LComm Bands (<=25%)	0	0	0	0	0	0	0	0	0	0	
No. LComm Bands (<=50%)	3	7	2	4	1	2	3	5	4	2	
Mean h	0,080	0,148	0,068	0,091	0,000	0,000	0,000	0,068	0,045	0,000	
SE of Mean h	0,054	0,063	0,046	0,061	0,000	0,000	0,000	0,046	0,045	0,000	
Mean uh	0,106	0,197	0,091	0,121	0,000	0,000	0,000	0,091	0,061	0,000	
SE of Mean uh	0,072	0,084	0,061	0,081	0,000	0,000	0,000	0,061	0,061	0,000	

No. Bands = No. of Different Bands

No. Bands Freq. $\geq 5\% =$ No. of Different Bands with a Frequency $\geq 5\%$

No. Private Bands = No. of Bands Unique to a Single Population

No. LComm Bands (<=25%) = No. of Locally Common Bands (Freq. >= 5%) Found in 25% or Fewer Populations

No. LComm Bands ($\leq 50\%$) = No. of Locally Common Bands (Freq. $\geq 5\%$) Found in 50% or Fewer Populations

 $h = Diversity = 1 - (p^2 + q^2)$

uh = Unbiased Diversity = (N / (N-1)) * h

Where for Haploid Binary data, p = Band Freq. and q = 1 - p.

			0	PB-01						
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
No. Bands	2	5	8	3	7	6	1	2	3	2
No. Bands Freq. >= 5%	2	5	8	3	7	6	1	2	3	2
No. Private Bands	0	0	0	0	1	0	0	0	0	0
No. LComm Bands (<=25%)	0	2	2	0	0	0	0	0	0	0
No. LComm Bands (<=50%)	1	4	7	1	4	4	0	0	1	0
Mean h	0,088	0,188	0,150	0,138	0,089	0,188	0,050	0,075	0,038	0,089
SE of Mean h	0,059	0,063	0,061	0,071	0,059	0,063	0,050	0,050	0,038	0,059
Mean uh	0,117	0,250	0,200	0,183	0,133	0,250	0,067	0,100	0,050	0,133
SE of Mean uh	0,079	0,083	0,082	0,094	0,089	0,083	0,067	0,067	0,050	0,089

			0	PB-04						
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
No. Bands	9	9	13	9	0	7	8	9	6	6
No. Bands Freq. >= 5%	9	9	13	9	0	7	8	9	6	6



No. Private Bands	0	0	1	0	0	0	0	0	0	1
No. LComm Bands (<=25%)	0	1	1	0	0	1	0	1	0	0
No. LComm Bands (<=50%)	7	7	9	7	0	6	6	7	4	3
Mean h	0,125	0,107	0,190	0,071	0,000	0,065	0,113	0,107	0,065	0,085
SE of Mean h	0,045	0,038	0,045	0,033	0,000	0,036	0,040	0,038	0,036	0,039
Mean uh	0,167	0,143	0,254	0,095	0,000	0,087	0,151	0,143	0,087	0,127
SE of Mean uh	0,060	0,051	0,060	0,044	0,000	0,048	0,054	0,051	0,048	0,059

OPG-02										
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
No. Bands	10	7	18	8	11	13	10	8	8	7
No. Bands Freq. >= 5%	10	7	18	8	11	13	10	8	8	7
No. Private Bands	0	0	1	0	0	1	0	0	0	0
No. LComm Bands (<=25%)	2	1	3	1	1	1	2	0	1	0
No. LComm Bands (<=50%)	8	6	13	4	8	9	7	4	5	4
Mean h	0,046	0,014	0,190	0,014	0,016	0,171	0,088	0,046	0,060	0,049
SE of Mean h	0,026	0,014	0,042	0,014	0,016	0,038	0,033	0,026	0,029	0,027
Mean uh	0,062	0,019	0,253	0,019	0,025	0,228	0,117	0,062	0,080	0,074
SE of Mean uh	0,035	0,019	0,057	0,019	0,025	0,050	0,043	0,035	0,038	0,041

OPZ-03												
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10		
No. Bands	9	6	9	14	11	11	13	11	12	9		
No. Bands Freq. >= 5%	9	6	9	14	11	11	13	11	12	9		
No. Private Bands	0	0	0	1	0	1	2	0	0	0		
No. LComm Bands (<=25%)	0	0	0	1	0	1	0	0	0	0		
No. LComm Bands (<=50%)	4	1	0	2	2	2	4	3	4	2		
Mean h	0,185	0,107	0,179	0,208	0,148	0,077	0,173	0,131	0,089	0,063		
SE of Mean h	0,049	0,038	0,047	0,045	0,047	0,036	0,045	0,042	0,036	0,035		
Mean uh	0,246	0,143	0,238	0,278	0,222	0,103	0,230	0,175	0,119	0,095		
SE of Mean uh	0,065	0,051	0,063	0,060	0,070	0,048	0,060	0,056	0,048	0,052		

			(C B-09						
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
No. Bands	11	13	12	13	9	9	13	11	12	15
No. Bands Freq. >= 5%	11	13	12	13	9	9	13	11	12	15
No. Private Bands	1	0	0	0	0	0	0	0	0	0



No. LComm Bands (<=25%)	1	1	0	1	0	0	1	0	1	1
No. LComm Bands (<=50%)	2	3	2	4	2	1	5	3	5	6
Mean h	0,033	0,109	0,065	0,147	0,019	0,130	0,130	0,098	0,196	0,174
SE of Mean h	0,023	0,040	0,036	0,044	0,019	0,043	0,038	0,035	0,048	0,046
Mean uh	0,043	0,145	0,087	0,196	0,029	0,174	0,174	0,130	0,261	0,261
SE of Mean uh	0,030	0,053	0,048	0,058	0,029	0,057	0,051	0,047	0,064	0,069

CB-12 Population P1 P2 P3 P4 P5 P6 P7 P8 P9 P10 No. Bands 14 16 13 16 16 14 15 17 19 10 No. Bands Freq. >= 14 16 13 16 16 14 15 17 19 10										
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
No. Bands	14	16	13	16	16	14	15	17	19	10
No. Bands Freq. >= 5%	14	16	13	16	16	14	15	17	19	10
No. Private Bands	1	0	0	0	0	0	1	0	0	0
No. LComm Bands (<=25%)	0	1	1	0	0	0	0	0	0	0
No. LComm Bands (<=50%)	2	5	5	3	5	4	3	6	7	2
Mean h	0,098	0,138	0,031	0,161	0,222	0,165	0,156	0,143	0,223	0,063
SE of Mean h	0,033	0,039	0,022	0,039	0,043	0,043	0,038	0,041	0,041	0,030
Mean uh	0,131	0,185	0,042	0,214	0,333	0,220	0,208	0,190	0,298	0,095
SE of Mean uh	0,044	0,052	0,029	0,052	0,064	0,058	0,050	0,054	0,054	0,045

			(C B-13						
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
No. Bands	10	22	16	19	14	18	14	18	13	15
No. Bands Freq. >= 5%	10	22	16	19	14	18	14	18	13	15
No. Private Bands	1	3	0	2	0	0	0	0	0	1
No. LComm Bands (<=25%)	1	1	3	2	0	3	1	4	1	2
No. LComm Bands (<=50%)	7	15	12	14	12	13	10	14	11	12
Mean h	0,070	0,185	0,096	0,104	0,083	0,151	0,117	0,156	0,065	0,120
SE of Mean h	0,023	0,031	0,024	0,027	0,025	0,029	0,027	0,031	0,023	0,029
Mean uh	0,094	0,247	0,128	0,139	0,125	0,201	0,156	0,208	0,087	0,181
SE of Mean uh	0,031	0,042	0,033	0,036	0,038	0,038	0,036	0,042	0,031	0,043

			(C B-15						
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
No. Bands	11	6	12	9	6	13	15	12	14	8
No. Bands Freq. >= 5%	11	6	12	9	6	13	15	12	14	8
No. Private Bands	1	0	1	0	0	0	1	1	1	0
No. LComm Bands (<=25%)	0	1	0	2	0	2	1	1	1	0



No. LComm Bands (<=50%)	2	1	6	3	1	7	7	5	6	3
Mean h	0,154	0,091	0,154	0,144	0,085	0,149	0,159	0,106	0,154	0,017
SE of Mean h	0,039	0,034	0,039	0,040	0,035	0,038	0,041	0,035	0,043	0,017
Mean uh	0,205	0,122	0,205	0,192	0,128	0,199	0,212	0,141	0,205	0,026
SE of Mean uh	0,053	0,045	0,053	0,054	0,053	0,051	0,054	0,047	0,057	0,026

	ALL MARKERS											
Population	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10		
No. Bands	80	92	104	97	76	93	93	93	91	75		
No. Bands Freq. >= 5%	80	92	104	97	76	93	93	93	91	75		
No. Private Bands	4	3	3	4	1	2	4	1	1	2		
No. LComm Bands (<=25%)	4	8	10	7	1	8	5	6	4	3		
No. LComm Bands (<=50%)	36	49	56	42	35	48	45	47	47	34		
Mean h	0,095	0,121	0,122	0,117	0,081	0,131	0,121	0,113	0,110	0,081		
SE of Mean h	0,012	0,013	0,013	0,013	0,012	0,013	0,013	0,013	0,013	0,012		
Mean uh	0,126	0,161	0,162	0,157	0,121	0,175	0,161	0,150	0,147	0,121		
SE of Mean uh	0,016	0,017	0,017	0,017	0,018	0,018	0,017	0,017	0,017	0,018		



Nei's Genetic Distance and Genetic Identity

OPA-04

		Pairw	ise Popul	ation Mat	trix of Nei	i Genetic I	Distance			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
0,000										P1
0,485	0,000									P2
0,273	0,491	0,000								P3
0,651	0,479	0,267	0,000							P4
0,309	0,569	0,011	0,271	0,000						P5
0,607	0,569	0,417	0,558	0,452	0,000					P6
0,132	0,660	0,417	0,964	0,452	0,788	0,000				P7
0,763	0,392	0,823	1,216	0,859	0,348	0,491	0,000			P8
0,584	0,503	0,635	1,411	0,670	0,503	0,360	0,062	0,000		P9
0,187	0,485	0,283	0,404	0,318	0,318	0,318	0,753	0,871	0,000	P10

		Pairw	vise Popul	ation Ma	trix of Ne	i Genetic	Identity			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
1,000										P1
0,616	1,000									P2
0,761	0,612	1,000								P3
0,522	0,620	0,765	1,000							P4
0,734	0,566	0,989	0,763	1,000						P5
0,545	0,566	0,659	0,572	0,636	1,000					P6
0,876	0,517	0,659	0,381	0,636	0,455	1,000				P7
0,466	0,676	0,439	0,296	0,424	0,706	0,612	1,000			P8
0,558	0,605	0,530	0,244	0,512	0,605	0,698	0,940	1,000		P9
0,829	0,615	0,753	0,667	0,727	0,727	0,727	0,471	0,419	1,000	P10



		Pairw	ise Popul	ation Mat	trix of Nei	i Genetic	Distance			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
0,000										P1
0,243	0,000									P2
0,644	0,326	0,000								P3
0,073	0,197	0,761	0,000							P4
0,688	1,141	0,748	0,624	0,000						P5
0,341	0,537	0,413	0,312	0,243	0,000					P6
0,007	0,227	0,692	0,063	0,726	0,381	0,000				P7
0,078	0,307	0,796	0,035	0,512	0,232	0,069	0,000			P8
0,158	0,327	0,848	0,040	0,488	0,234	0,148	0,020	0,000		P9
0,060	0,293	0,788	0,032	0,536	0,243	0,052	0,001	0,030	0,000	P10

		Pairv	vise Popul	lation Ma	trix of Ne	i Genetic	Identity			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
1,000										P1
0,784	1,000									P2
0,525	0,722	1,000								P3
0,930	0,821	0,467	1,000							P4
0,503	0,320	0,473	0,536	1,000						P5
0,711	0,585	0,662	0,732	0,785	1,000					P6
0,993	0,797	0,501	0,939	0,484	0,683	1,000				P7
0,925	0,735	0,451	0,966	0,599	0,793	0,933	1,000			P8
0,854	0,721	0,428	0,960	0,614	0,792	0,863	0,980	1,000		P9
0,941	0,746	0,455	0,968	0,585	0,785	0,949	0,999	0,970	1,000	P10

OPB-01



		Pairw	vise Popul	ation Mat	trix of Nei	i Genetic	Distance			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
0,000										P1
0,366	0,000									P2
0,317	0,397	0,000								P3
0,302	0,060	0,481	0,000							P4
0,253	0,314	0,265	0,299	0,000						P5
0,617	0,534	0,330	0,646	0,286	0,000					P6
0,343	0,443	0,258	0,422	0,196	0,312	0,000				P7
0,405	0,416	0,605	0,339	0,247	0,315	0,443	0,000			P8
0,341	0,389	0,379	0,353	0,223	0,412	0,425	0,143	0,000		P9
0,325	0,577	0,410	0,550	0,167	0,447	0,307	0,353	0,253	0,000	P10

		Pairv	vise Popul	lation Ma	trix of Ne	i Genetic	Identity			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
1,000										P1
0,694	1,000									P2
0,728	0,672	1,000								P3
0,740	0,941	0,618	1,000							P4
0,776	0,731	0,767	0,741	1,000						P5
0,540	0,586	0,719	0,524	0,751	1,000					P6
0,709	0,642	0,773	0,656	0,822	0,732	1,000				P7
0,667	0,660	0,546	0,713	0,781	0,730	0,642	1,000			P8
0,711	0,678	0,684	0,703	0,800	0,662	0,654	0,867	1,000		P9
0,723	0,562	0,664	0,577	0,846	0,639	0,736	0,702	0,777	1,000	P10

OPB-04



		Pairw	vise Popul	ation Mat	trix of Ne	i Genetic	Distance			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
0,000										P1
0,447	0,000									P2
0,583	0,658	0,000								P3
0,447	0,449	0,492	0,000							P4
0,534	0,672	0,072	0,567	0,000						P5
0,330	0,362	0,332	0,521	0,336	0,000					P6
0,249	0,543	0,297	0,352	0,319	0,272	0,000				P7
0,358	0,541	0,510	0,098	0,534	0,470	0,282	0,000			P8
0,358	0,454	0,548	0,144	0,577	0,523	0,301	0,063	0,000		P9
0,495	0,475	0,509	0,082	0,577	0,497	0,284	0,104	0,086	0,000	P10

		Pairv	vise Popul	lation Ma	trix of Ne	i Genetic	Identity			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
1,000										P1
0,640	1,000									P2
0,558	0,518	1,000								P3
0,640	0,638	0,611	1,000							P4
0,586	0,511	0,930	0,567	1,000						P5
0,719	0,697	0,718	0,594	0,715	1,000					P6
0,779	0,581	0,743	0,703	0,727	0,762	1,000				P7
0,699	0,582	0,600	0,907	0,586	0,625	0,755	1,000			P8
0,699	0,635	0,578	0,866	0,562	0,593	0,740	0,939	1,000		P9
0,609	0,622	0,601	0,921	0,562	0,609	0,752	0,901	0,918	1,000	P10

OPG-02



	Pairwise Population Matrix of Nei Genetic Distance												
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10				
0,000										P1			
0,129	0,000									P2			
0,205	0,056	0,000								P3			
0,341	0,297	0,236	0,000							P4			
0,442	0,330	0,158	0,239	0,000						P5			
0,417	0,442	0,331	0,303	0,146	0,000					P6			
0,182	0,388	0,431	0,369	0,449	0,446	0,000				P7			
0,347	0,412	0,360	0,109	0,409	0,548	0,305	0,000			P8			
0,370	0,456	0,436	0,181	0,468	0,630	0,282	0,058	0,000		P9			
0,667	0,443	0,298	0,149	0,246	0,494	0,606	0,179	0,240	0,000	P10			

		Pairv	vise Popul	lation Ma	trix of Ne	i Genetic	Identity			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
1,000										P1
0,879	1,000									P2
0,815	0,945	1,000								P3
0,711	0,743	0,790	1,000							P4
0,643	0,719	0,854	0,788	1,000						P5
0,659	0,643	0,718	0,738	0,864	1,000					P6
0,833	0,679	0,650	0,691	0,638	0,640	1,000				P7
0,707	0,662	0,697	0,897	0,664	0,578	0,737	1,000			P8
0,691	0,634	0,647	0,834	0,626	0,532	0,754	0,943	1,000		P9
0,513	0,642	0,742	0,862	0,782	0,610	0,545	0,836	0,786	1,000	P10

OPZ-03



		Pairw	vise Popul	ation Ma	trix of Nei	i Genetic	Distance			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
0,000										P1
0,496	0,000									P2
0,520	0,370	0,000								P3
0,544	0,188	0,438	0,000							P4
0,570	0,453	0,174	0,488	0,000						P5
0,410	0,351	0,184	0,330	0,103	0,000					P6
0,410	0,405	0,526	0,522	0,441	0,412	0,000				P7
0,816	0,675	0,376	0,562	0,484	0,440	0,540	0,000			P8
0,720	0,718	0,529	0,561	0,650	0,441	0,548	0,113	0,000		P9
0,552	0,540	0,564	0,633	0,910	0,668	0,528	0,314	0,138	0,000	P10

		Pairw	vise Popul	lation Ma	trix of Ne	i Genetic	Identity			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
1,000										P1
0,609	1,000									P2
0,594	0,691	1,000								P3
0,580	0,829	0,645	1,000							P4
0,565	0,636	0,840	0,614	1,000						P5
0,664	0,704	0,832	0,719	0,902	1,000					P6
0,664	0,667	0,591	0,593	0,643	0,663	1,000				P7
0,442	0,509	0,686	0,570	0,616	0,644	0,583	1,000			P8
0,487	0,488	0,589	0,571	0,522	0,643	0,578	0,893	1,000		P9
0,576	0,583	0,569	0,531	0,403	0,513	0,590	0,730	0,871	1,000	P10



		Pairw	vise Popul	ation Mat	trix of Ne	i Genetic	Distance			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
0,000										P1
0,192	0,000									P2
0,532	0,454	0,000								P3
0,375	0,359	0,381	0,000							P4
0,382	0,407	0,287	0,411	0,000						P5
0,364	0,334	0,269	0,374	0,194	0,000					P6
0,219	0,377	0,483	0,290	0,344	0,339	0,000				P7
0,529	0,456	0,515	0,172	0,455	0,562	0,273	0,000			P8
0,463	0,596	0,792	0,323	0,297	0,532	0,372	0,310	0,000		P9
0,312	0,458	0,863	0,388	0,277	0,381	0,406	0,560	0,145	0,000	P10

		Pairv	vise Popul	lation Ma	trix of Ne	i Genetic	Identity			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
1,000										P1
0,826	1,000									P2
0,587	0,635	1,000								P3
0,688	0,698	0,683	1,000							P4
0,682	0,665	0,751	0,663	1,000						P5
0,695	0,716	0,764	0,688	0,824	1,000					P6
0,804	0,686	0,617	0,748	0,709	0,713	1,000				P7
0,589	0,634	0,598	0,842	0,634	0,570	0,761	1,000			P8
0,629	0,551	0,453	0,724	0,743	0,588	0,689	0,733	1,000		P9
0,732	0,633	0,422	0,678	0,758	0,683	0,666	0,571	0,865	1,000	P10



		Pairw	ise Popul	ation Ma	trix of Nei	i Genetic	Distance			
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	
0,000										P1
0,159	0,000									P2
0,261	0,222	0,000								P3
0,338	0,280	0,328	0,000							P4
0,302	0,322	0,243	0,344	0,000						P5
0,105	0,111	0,138	0,273	0,195	0,000					P6
0,192	0,148	0,175	0,284	0,170	0,113	0,000				P7
0,194	0,214	0,177	0,220	0,231	0,110	0,161	0,000			P8
0,282	0,314	0,386	0,357	0,265	0,226	0,271	0,234	0,000		P9
0,232	0,226	0,293	0,345	0,257	0,146	0,187	0,172	0,103	0,000	P10

	Pairwise Population Matrix of Nei Genetic Identity											
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	_		
1,000										P1		
0,853	1,000									P2		
0,770	0,801	1,000								P3		
0,713	0,756	0,721	1,000							P4		
0,739	0,725	0,784	0,709	1,000						P5		
0,900	0,895	0,871	0,761	0,823	1,000					P6		
0,825	0,863	0,840	0,753	0,843	0,893	1,000				P7		
0,823	0,807	0,838	0,803	0,794	0,895	0,851	1,000			P8		
0,754	0,731	0,680	0,700	0,767	0,798	0,762	0,792	1,000		P9		
0,793	0,798	0,746	0,708	0,773	0,864	0,829	0,842	0,902	1,000	P10		



	Pairwise Population Matrix of Nei Genetic Distance										
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10		
0,000										P1	
0,150	0,000									P2	
0,215	0,150	0,000								P3	
0,133	0,030	0,133	0,000							P4	
0,143	0,071	0,109	0,060	0,000						P5	
0,298	0,225	0,073	0,202	0,193	0,000					P6	
0,233	0,416	0,355	0,403	0,360	0,243	0,000				P7	
0,300	0,382	0,462	0,400	0,461	0,326	0,283	0,000			P8	
0,383	0,362	0,443	0,372	0,433	0,329	0,347	0,074	0,000		P9	
0,323	0,316	0,447	0,348	0,391	0,433	0,396	0,190	0,129	0,000	P10	

	Pairwise Population Matrix of Nei Genetic Identity										
P1	P2	P3	P4	P5	P6	P7	P8	P9	P10		
1,000										P1	
0,861	1,000									P2	
0,807	0,861	1,000								P3	
0,876	0,970	0,876	1,000							P4	
0,867	0,932	0,896	0,942	1,000						P5	
0,742	0,798	0,929	0,817	0,825	1,000					P6	
0,792	0,660	0,701	0,669	0,698	0,784	1,000				P7	
0,741	0,683	0,630	0,670	0,631	0,722	0,754	1,000			P8	
0,682	0,696	0,642	0,689	0,649	0,720	0,707	0,929	1,000		P9	
0,724	0,729	0,640	0,706	0,676	0,648	0,673	0,827	0,879	1,000	P10	